



BML486056

44828

2411496773

07/11/2024 13:00

07/11/2024 17:23

: 07/11/2024 16:11

Laboratory Investigation Report

Name : Ms. CHARIZE ALEJANDRO

DOB : 03/10/1990

Age / Gender : 34 Y 1 M / Female

Referred by : Dr. Enomen Goodluck Ekata
Centre : CITICARE MEDICAL CENTER

BIOCHEMISTRY

Result Flag Unit Test Reference Range Methodology **AMYLASE TOTAL** 76 U/L 28 - 100 Enzymatic colorimetric assay acc to IFCC Please note change. Source: Roche IFU. Particle-enhanced **C-REACTIVE PROTEIN (CRP)** < 5.0 17 mg/L

Please note change.

Source: Roche IFU.

note change immunoturbidimetric assay

Ref No.

Sample No.

Collected

Registered

Reported

Comments: Please correlate clinically.

INTERPRETATION NOTES:

- 1. CRP measurements are used as aid in diagnosis, monitoring, prognosis, and management of suspected inflammatory disorders and associated diseases, acute infections and tissue injury.
- 2. C-reactive protein is the classic acute phase protein in inflammatory reactions.
- 3. CRP is the most sensitive of the acute phase reactants and its concentration increases rapidly during inflammatory processes. The CRP response frequently precedes clinical symptoms, including fever. After onset of an acute phase response, the serum CRP concentration rises rapidly and extensively. The increase begins within 6 to 12 hours and the peak value is reached within 24 to 48 hours. Levels above 100 mg/L are associated with severe stimuli such as major trauma and severe infection (sepsis).
- 1. CRP response may be less pronounced in patients suffering from liver disease.
- 5. CRP assays are used to detect systemic inflammatory processes (apart from certain types of inflammation such as systemic lupus erythematosus (SLE) and Colitis ulcerosa); to assess treatment of bacterial infections with antibiotics; to detect intrauterine infections with concomitant premature amniorrhexis; to differentiate between active and inactive forms of disease with concurrent infection, e.g. in patients suffering from SLE or Colitis ulcerosa; to therapeutically monitor rheumatic disease and assess anti-inflammatory therapy; to determine the presence of post-operative complications at an early stage, such as infected wounds, thrombosis and pneumonia, and to distinguish between infection and bone marrow transplant rejection.

LIPASE 69 H U/L 13 - 60 Enzymatic colorimetric assay

Please note change. Source: Roche IFU.

Dr. Adley Mark Fernandes

Dr. Vyoma V Shah

M.D (Pathology)

M.D (Pathology)

This is an electronically authenticated report

P.O Box: 49527

Pathologist

an

MOHAMMED RASHID CHENANGADATH

Laboratory Technologist
Printed on: 07/11/2024 19:02

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Dubai, UAE

Clinical Pathologist



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BIOCHEMISTRY

Result Flag Unit **Reference Range** Test

Methodology

INTERPRETATION NOTES:

- 1. Lipases are group of enzymes which catalyze the cleavage of triglycerides to diglycerides with subsequent formation of monoglycerides and fatty acids.
- 2. Lipase is produced by the pancreas, liver, intestine, tongue, stomach, and many other cells.
- 3. The lipase activity determination has gained increasing international recognition because of its high specificity and rapid response. After acute pancreatitis, the lipase activity increases within 4-8 hours, reaches a peak after 24 hours and decreases after 8 to 14 days. However, there is no correlation between the lipase activity determined in serum and the extent of damage to the pancreas.
- 4. Because lipase levels remain elevated longer than amylase and its sensitivity in acute alcoholic pancreatitis is increased, serum lipase may be a more reliable test than serum amylase for the initial diagnosis of acute pancreatitis. Daily measurements of lipase are of no value in the assessment of the patient's clinical progress or ultimate prognosis. Because of its sensitivity, lipase testing is not very useful in chronic pancreatitis or pancreatic cancer.
- 5. Along with pancreatic disorders, lipase testing is also indicated in the diagnosis of peritonitis, strangulated or infarcted bowel, and pancreatic cyst.

References:

1. Kit insert

2. Williamson MA, Snyder LM, Wallach JB. Wallach's interpretation of diagnostic tests. 9th ed. Philadelphia: Wolters Kluwer/Lippincott Williams & Wilkins Health; 2011.

Sample Type: Serum

End of Report

Dr. Adley Mark Fernandes M.D (Pathology) **Pathologist**

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Dr. Vyoma V Shah M.D (Pathology) **Clinical Pathologist**

Gome V. Shah

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Centre : CITICARE MEDICAL CENTER

| HE | MA [*] | TOL | .OGY |
|----|-----------------|-----|------|

| Test | Result | Flag | Unit | Reference Range | Methodology | | |
|--------------------------------|--------|------|----------------|-----------------|----------------------|--|--|
| COMPLETE BLOOD COUNT (CBC) | | | | | | | |
| HEMOGLOBIN | 13.8 | | g/dL | 12 - 15.5 | Photometric | | |
| RBC COUNT | 4.8 | | 10^6/μL | 3.9 - 5 | Electrical Impedance | | |
| HEMATOCRIT | 42.1 | | % | 35 - 45 | Calculation | | |
| MCV | 88.3 | | fL | 82 - 98 | Calculation | | |
| мсн | 28.8 | | pg | 27 - 32 | Calculation | | |
| мснс | 32.7 | | g/dL | 32 - 37 | Calculation | | |
| RDW | 14.3 | | % | 11.9 - 15.5 | Calculation | | |
| RDW-SD | 43.8 | | fL | | Calculation | | |
| MPV | 8.2 | | fL | 7.6 - 10.8 | Calculation | | |
| PLATELET COUNT | 332 | | 10^3/uL | 150 - 450 | Electrical Impedance | | |
| РСТ | 0.3 | | % | 0.01 - 9.99 | Calculation | | |
| PDW | 16.1 | | Not Applicable | 0.1 - 99.9 | Calculation | | |
| NUCLEATED RBC (NRBC)^ | 0.1 | | /100 WBC | | VCS 360 Technology | | |
| ABSOLUTE NRBC COUNT^ | 0.02 | | 10^3/uL | | Calculation | | |
| EARLY GRANULOCYTE COUNT (EGC)^ | 0.4 | | % | | VCS 360 Technology | | |
| ABSOLUTE EGC^ | 0.1 | | 10^3/uL | | Calculation | | |
| WBC COUNT | 14.1 | н | 10^3/μL | 4 - 11 | Electrical Impedance | | |
| DIFFERENTIAL COUNT (DC) | | | | | | | |
| NEUTROPHILS | 65 | | % | 40 - 75 | VCS 360 Technology | | |
| LYMPHOCYTES | 27 | L | % | 30 - 60 | VCS 360 Technology | | |
| EOSINOPHILS | 3 | | % | 0 - 6 | VCS 360 Technology | | |
| MONOCYTES | 5 | | % | 1 - 6 | VCS 360 Technology | | |
| BASOPHILS | 0 | | % | 0 - 1 | VCS 360 Technology | | |
| ABSOLUTE COUNT | | | | | | | |
| ABSOLUTE NEUTROPHIL COUNT | 9.1 | н | 10^3/uL | 1.6 - 8.25 | Calculation | | |
| ABSOLUTE LYMPHOCYTE COUNT | 3.8 | | 10^3/uL | 1.2 - 6.6 | Calculation | | |
| ABSOLUTE MONOCYTE COUNT | 0.7 | н | 10^3/uL | 0.04 - 0.66 | Calculation | | |
| ABSOLUTE EOSINOPHIL COUNT | 0.4 | | 10^3/uL | 0 - 0.66 | Calculation | | |
| ABSOLUTE BASOPHIL COUNT | 0.0 | | 10^3/uL | 0 - 0.11 | Calculation | | |

Comments : Please correlate clinically.

Dr. Adley Mark Fernandes

M.D (Pathology)

Pathologist

Dr. Vyoma V Shah

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Clinical Pathologist

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Thahsina Anees
Laboratory Technologist

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HEMATOLOGY

End of Report

Test Result Flag Unit Reference Range Methodology

COMPLETE BLOOD COUNT (CBC)

INTERPRETATION NOTES:

Please note update on CBC report format, reference ranges and method(Beckman Coulter).

Sample Type: EDTA Whole Blood

Dr. Adley Mark Fernandes M.D (Pathology) Pathologist

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Dr. Vyoma V Shah M.D (Pathology) Clinical Pathologist

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Thahsina Anees
Laboratory Technologist

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SEROLOGY

Test Result Flag Unit Reference Range Methodology
HELICOBACTER PYLORI ANTIBODY Negative Negative Immunochromatography

Sample Type : Serum

End of Report



Dr. Adley Mark Fernandes M.D (Pathology) Pathologist

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Dr. Vyoma V Shah M.D (Pathology) Clinical Pathologist

Gome V. Shah

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NAZAR MOHAMED ALI Laboratory Technologist Printed on: 07/11/2024 19:02

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